

**Professional  
Practice**  
*in Clinical Chemistry*

# Method Validation

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**Professional  
Practice**  
*in Clinical Chemistry*

# Learning Objectives

**After this presentation, you should be able to:**

1. Define method evaluation.
2. List the steps needed to complete a method evaluation study.
3. Define total allowable error (TEa).
4. Apply TEa to method evaluation.
5. Describe recommendations for Sigma values.



# Looking to implement a clinical test?

- Establish the need
- Clinical performance
  - Clinical sensitivity
  - Clinical specificity
- Define the performance standards
  - Costs/efficiencies/space
  - Turn around times/sample requirements
  - Analytical Quality (from kit insert, references)
- Select the new method
- Evaluate the new method
- Implement the new method



# What is method evaluation?

- Determination of:
  - analytical performance characteristics
  - clinical performance characteristics
- Validation
  - Objective evidence that requirements for a specific intended use can be fulfilled consistently
- Verification
  - Objective evidence that requirements have been fulfilled



# What do you do?

- FDA approved?
  - Clinical Laboratory Improvement Amendments (CLIA) requirements
  - Match performance specs established by the manufacturer
    - Accuracy | Should be comparable to manufacture's
    - Precision | Should be smaller than CLIA requirement
    - Reportable Range | Appropriate for patient care
    - Verify manufacturer's reference intervals
    - Determine test system calibration and control procedures based on specs above
    - Document all activities



# Experiments to Validate?

- FDA approved?
  - Reportable Range
    - Linearity
  - Precision
    - Within-run precision
    - Total precision and QC ranges
  - Accuracy
    - Comparison of methods
  - Reference Intervals



# Why?

- Clinical significance - leads to accurate medical decisions
- Required by CLIA\*, CAP, and The Joint Commission  
(\*Clinical Laboratory Improvements Amendments of 1988)
- Pass proficiency testing
- Improvements over existing methodology
- Assay validation requirements vary:  
Non-FDA approved > FDA approved > Waived tests  
Today we are going to focus on  
**FDA approved, non-waived tests**



# Steps in Method Validation

- 1) Define Goals
- 2) Error Assessment
- 3) Compare error vs. analytical goal





# 1<sup>st</sup> Step in Method Validation

## Define Goals

- Accept that all lab measurements contain experimental error
- What is an acceptable performance for:
  - Precision?
  - Accuracy?
  - Sensitivity?
  - Analytical measurement range?



# Define Goals

- Lab error should be:
  - smaller than CLIA (or other regulatory) requirement:
    - CLIA / 2?
    - CLIA / 3?
    - CLIA / 4?
    - CLIA / 6?
  - consistent with manufacturer's claims
  - compatible with patients' care



# 2<sup>nd</sup> Step in Method Validation

## Error Assessment

- Method validation assesses
  - Type of error
  - Magnitude of error
  - Clinical Significance of error
    - Literature guidelines
    - Physician input
    - Professional judgment



# 3<sup>rd</sup> Step in Method Validation

Compare error vs. analytical goal

Accept or reject your new method



# Accuracy and Precision

Accuracy – closeness of measured value to the “true” value – bias

Precision – dispersion of repeated measurements about the mean – reproducibility

Reliability –  
Accuracy + Precision

## Precision & Accuracy

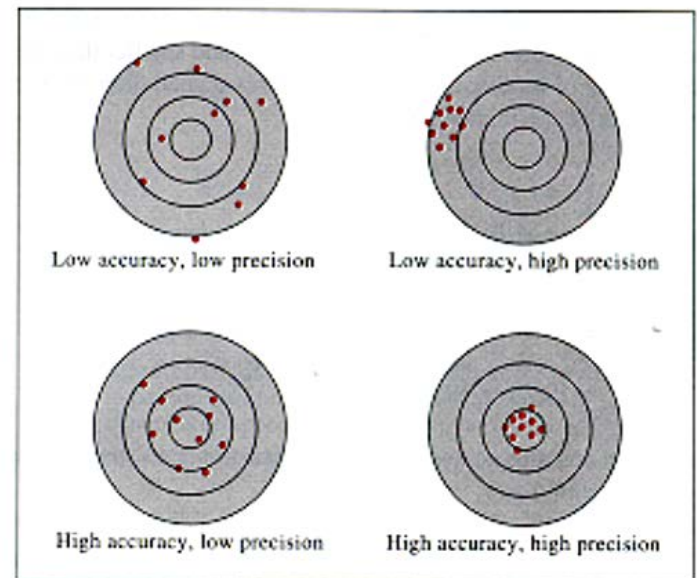
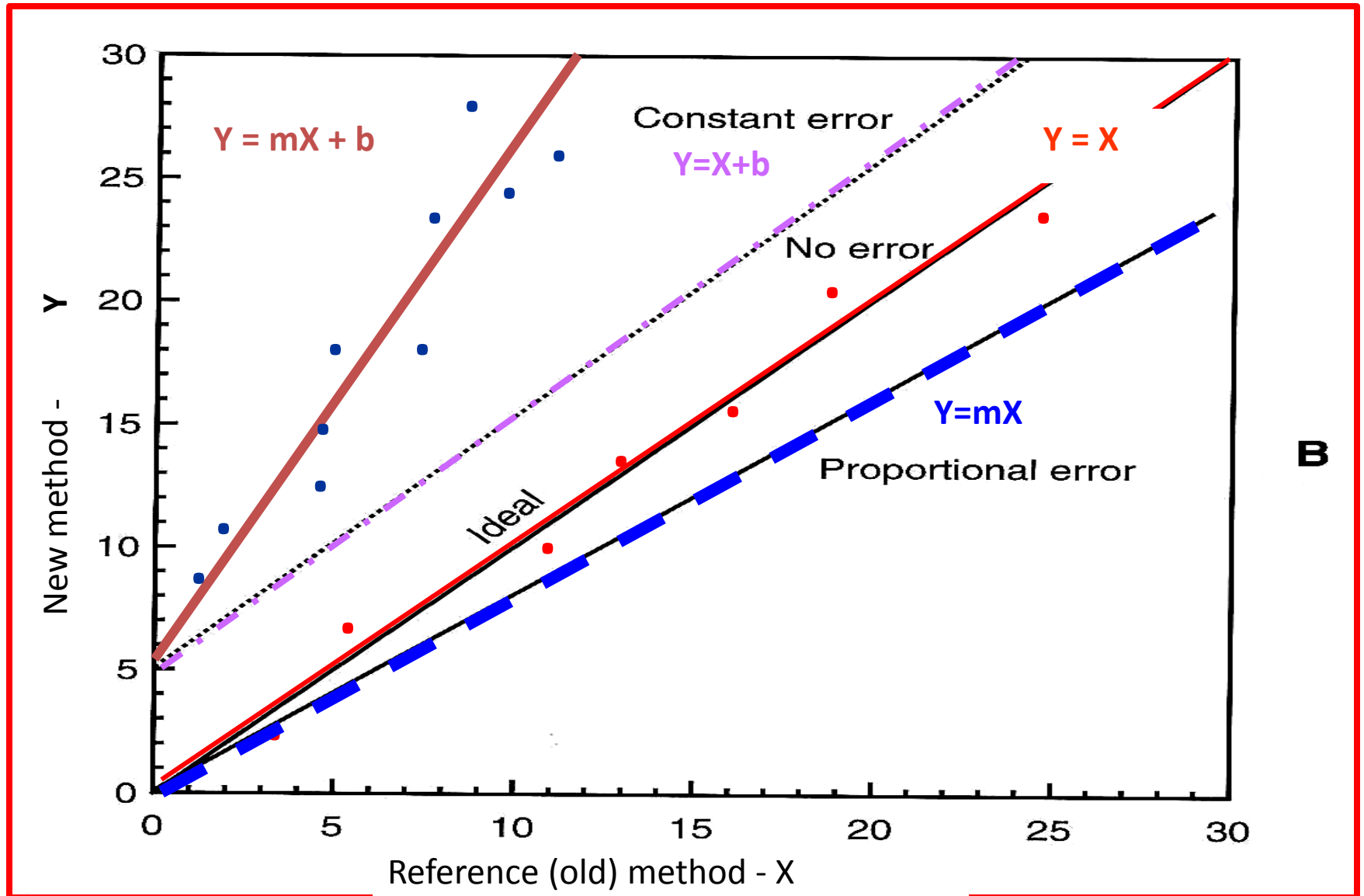


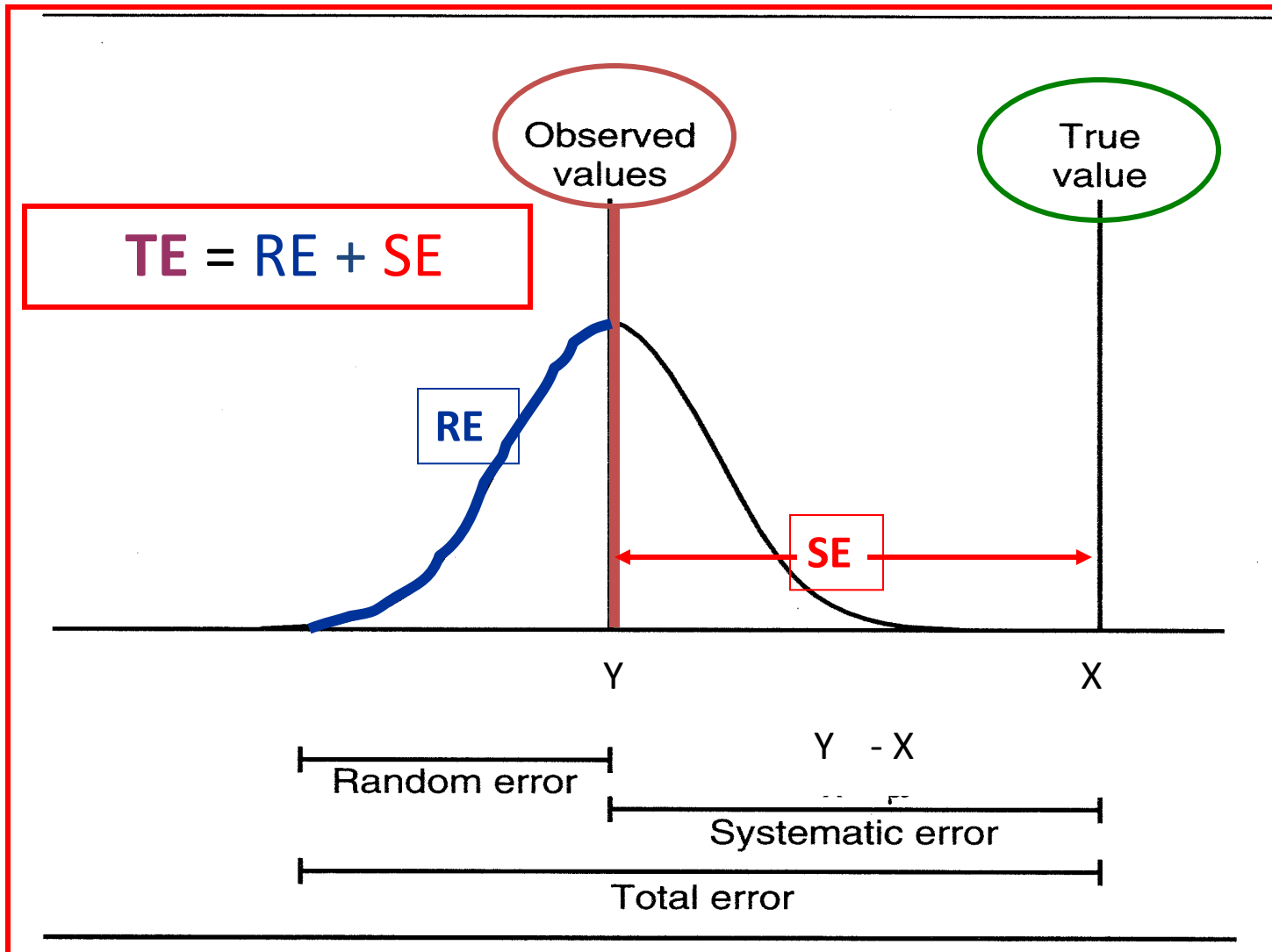
Figure 2-2  
Accuracy and precision.



# Systematic and Random Errors



# Total Analytical Error - TE



# Systematic Error - Affects accuracy

## Systematic error (SE) - Bias

- Types of systemic errors:
  - Proportional (indicated by slope)
  - Constant (indicated by intercept)
  - Proportional + Constant (Combination of both)
  - Caused by (examples): bad calibrators, bad reagents, bad pipettes, interference





# Random Error (RE) - Affects precision

- May be caused by (for example):
  - Variability in volume of sample or reagent delivered
  - Changes in environment
  - Inconsistent handling of materials
- Estimated by:
  - Standard deviation (SD)
  - Coefficient of variation (CV)
  - Correlation coefficient ( $r$ )



# Magnitude of Error – TE

- TE is the total maximum error of a test as measured in the lab
- TE is the sum of: random + systemic errors

$$\text{TE} = \text{RE} + \text{SE}$$

- Determined
  - For each given method
  - At various medical decision levels ( $X_c$ )



# Total Allowable Error - $TE_A$

$TE_A$  is the total error permitted by CLIA, based on

- Medical requirements
- Best available analytical method
- Compatible with proficiency testing expectations

**Goal:** Total Analytical Error < Total Allowable Error

$$TE < TE_A$$

Determined

- Method specific
- Measured at various Medical decision levels ( $X_c$ )



# Ready to Validate?

- FDA approved?
  - Reportable Range
    - Linearity
  - Precision
    - Within-run precision
    - Total precision and QC ranges
  - Accuracy
    - Comparison of methods
  - Reference Intervals



# AMR: Linearity Study

- Analytical Measurement Range (AMR)
  - Range of analyte where results are proportional to the true concentration of analyte in the sample
  - Range over which the test can be performed w/o modification (e.g. no dilution)
- Also called: Dynamic Range, and Reportable range
- Determined in the lab by linearity experiments



# AMR vs. MD/C

- **Analytical Measurement Range – AMR**
  - Range of analyte values that a method can directly measure w/o modification (no dilutions, concentrations, other pretreatments that are not part of the usual assay process)
- **Maximum Dilution/Concentration (formerly Clinically Reportable Range – CRR)**
  - Range of analyte values which are clinically significant
  - Can be reported following modification (such as dilutions)



# AMR vs. MD/C

Measurement range should be medically useful if:

- $MD/C > AMR$ 
  - Value higher than AMR: report as  $> X$  or dilute
  - Value lower than AMR : report as  $< X$  or concentrate

If:  $MD/C < AMR$  - Limit AMR



# Linearity Study – “to do” list

- Samples:
  - Ideal: Use “traceable” standards in matrix matched sample
  - Mix of very high with very low pt.’s samples are OK if conc. are known
  - Dilute high samples in acceptable matrix diluent
- At least 5-7 different conc. points within the reportable range (5 – 95% of AMR), equally spaced is ideal
- Testing is performed in duplicate
- Run from lowest to highest (to avoid carryover)
- Pipetting accuracy and precision is critical





# Limit of Detection

- Limit of Blank (LoB):
  - The lowest concentration that can be distinguished from background (blank, zero) noise
  - Sometimes called limit of absence.
  - Calculated as: Mean conc. of blank zero (>20 replicates) + 2SD
  - This is the number provided in most kit inserts
- Limit of Detection (LoD):
  - The lowest number that will almost always have a non-zero result (mean conc. of blank + 3 SD)
- ★ Limit of Quantification (LoQ):
  - The lowest concentration that can be quantified reliably
  - Analyte lowest concentration where  $CV \leq 20\%$  (or other error goal)
  - Results with higher CV% have large random error, thus are not useful for clinical interpretation

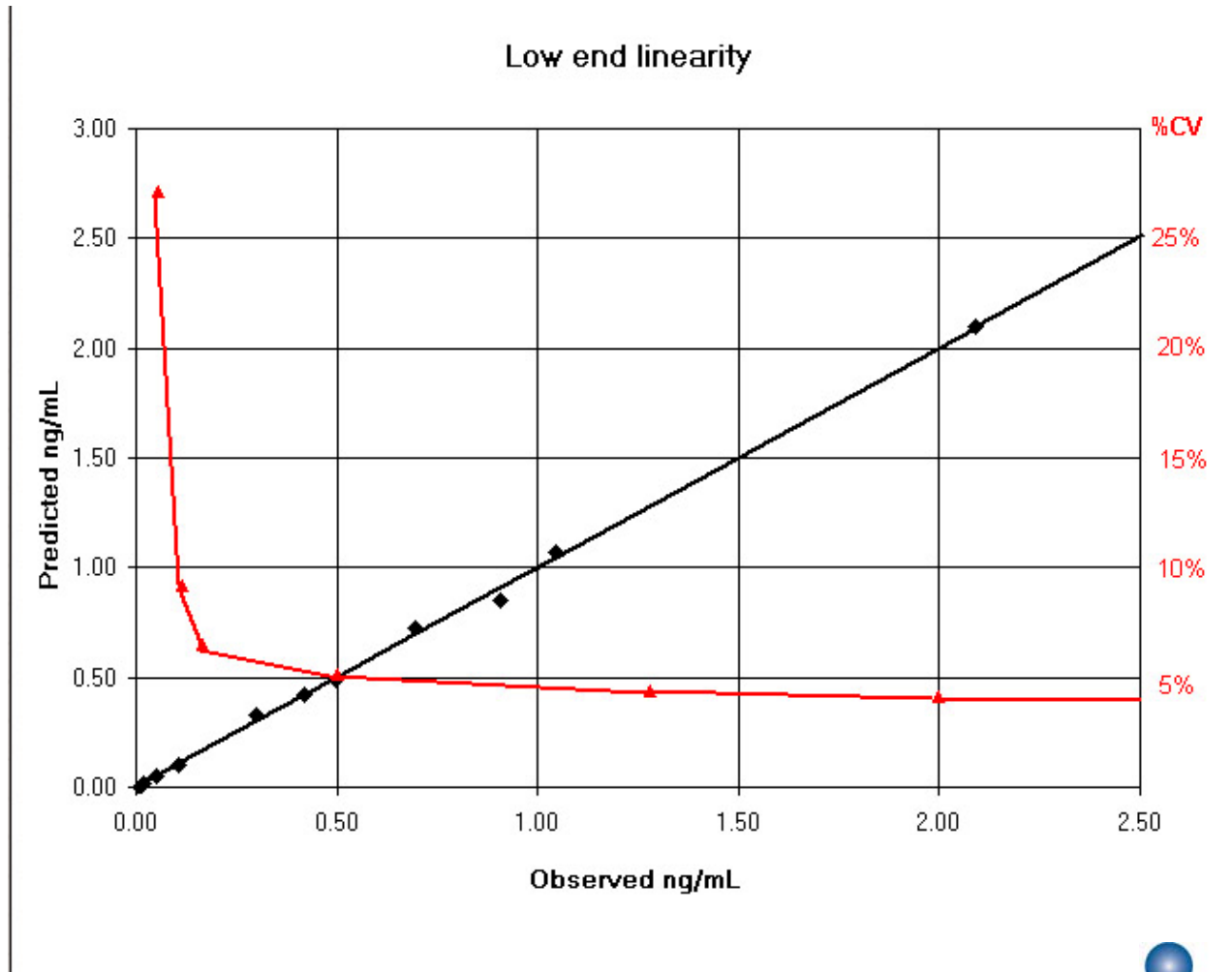


# LOQ Experiments

- Only needed if MD/C begins
  - At or near zero
  - At or below the manufacturer's stated AMR
  - Not necessary for most assays
- Start with low end linearity study
  - Determine the low end AMR
- Follow up with precision study
  - Calculate the precision (CV) at low end concentrations



# LOQ study example



# Experiments to Validate?

- FDA approved?
  - Reportable Range
    - Linearity
  - Precision
    - Within-run precision
    - Total precision and QC ranges
  - Accuracy
    - Comparison of methods
  - Reference Intervals



# Reproducibility Studies for Precision

## Random Error

- Use matrix matched samples
- Intra-Assay (within-run) Precision > 20x
- Inter-Assay (between-run) Precision > 20x
- Select specimens near medical decision levels
  - At least 2 control levels
- Calculate: mean, SD, CV%

Note: If you don't have established control limits, and they are being established during the experiment, revise limits every 5 days and look for evidence of unacceptable runs.

# Experiments to Validate?

- FDA approved?
  - Reportable Range
    - Linearity
  - Precision
    - Within-run precision
    - Total precision and QC ranges
  - Accuracy
    - Comparison of methods
  - Reference Intervals



# Method Comparison

## What do I do?

1. List results from two methods in pairs
  - Each pair represents the same sample
    - X – results of reference method
    - Y – results of new method
2. Create a scatter plot (plot the means of duplicates) if done in duplicate)
  - May also use a difference plot to analyze data
3. Look for outliers and data gaps
  - Repeat both methods for outliers
  - Try to fill in gaps or eliminate highest data during analysis



# Method Comparison

## What do I do?

4. Determine the correlation coefficient  
Check if “ $r$ ”  $> 0.975$

Note - Linear regression analysis may not be valid if the correlation coefficient is low.





# The correlation coefficient - r

- “r” – a statistical term
- It indicates the extent of linear relationship between the methods
- Ideally, r should be 1.00
- “r” can ranges from +1 to -1



# Characteristics of r

- “r” influenced by range of values
  - $r < 0.975$  may indicate that the range of data is too limited
- “r” is influenced by random errors only
- Systematic error has no effect on r
  - r is only used to assess linear relationship between methods
  - Method accuracy should not be based on r



# Method Comparison

## What do I do?

### 5. Generate a “linear best fit line”

$$Y = mX + b$$

m = slope (indicates a proportional error)

b = intercept (indicates constant error)



# Method Comparison

## What do I do?

### 6. Evaluate linear regression line:

Evaluate slope

Slope = 0.900 = -10% proportional error

Slope = 1.100 = +10% proportional error

Intercept should be close to zero (indicating very small constant bias)

May need to evaluate separate areas of the graph independently.



# Method Comparison

## What do I do?

### 7. Calculate systematic error at medical decision levels

Use slope and intercept to calculate systematic error:

$$Y_c = mX + b \quad SE = Y - X$$

$Y_c$  = Calculated result on new method

$X$  = Result from existing method

$m$  = Slope observed in method comparison experiment

$b$  = Intercept observed in method comparison experiment



# Method Comparison

## What do I do?

8. Compare result tracking over time. May be needed if:
  - Results are monitored over long intervals (trends)
  - The method comparison shows significant differences between the two methods



# Experiments to Validate?

- FDA approved?
  - Reportable Range
    - Linearity
  - Precision
    - Within-run precision
    - Total precision and QC ranges
  - Accuracy
    - Comparison of methods
  - Reference Intervals
    - Normal Range

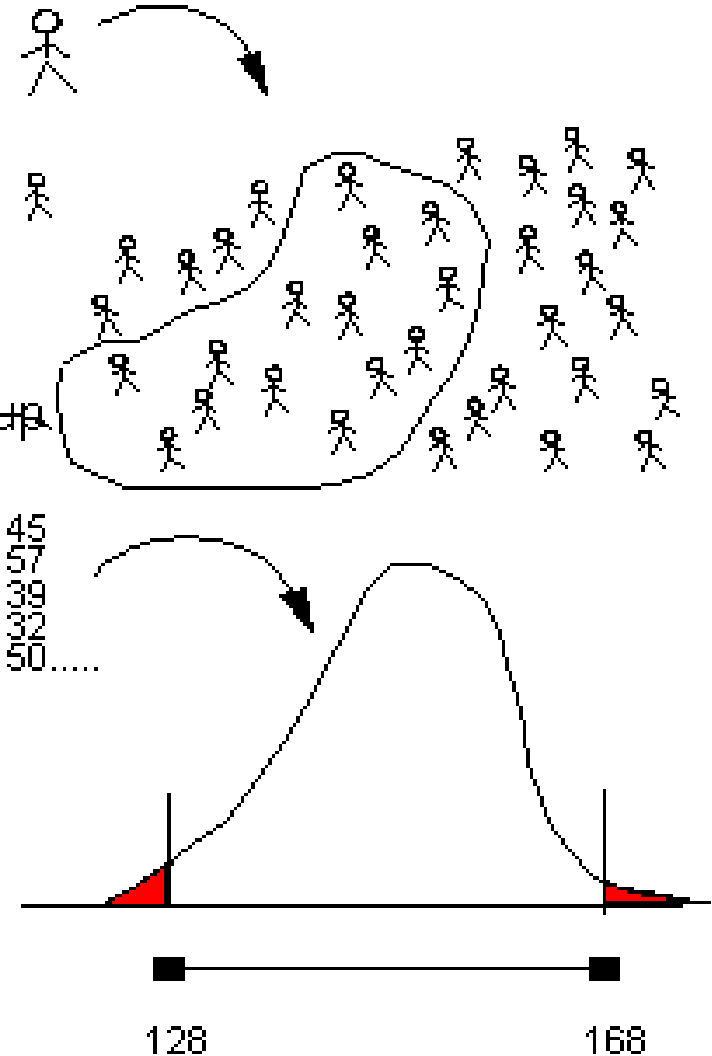


# The concept of reference values as recommended by the IFCC



## Reference Interval

reference individuals constitute a reference population from which is selected a reference sample group on which are determined reference values on which is observed a reference distribution on which is calculated reference limits that define a reference interval





# Reference Range Studies

- CLIA '88 requires verification of FDA approved manufacture's reference range
- Reference range study should reflect the laboratory's patient population
- Reference interval itself doesn't enter into the decision on method acceptability
- Usually done last, but testing should be done over several days.
- Data analysis will depend upon the distribution of the results.



# Reference Range Studies

- Validating a reference range: The number of samples needed if age/sex not a factor:
  - Verification of manufacturer's range  $N \geq 20$ 
    - Used if using the manufacturer's range and the test will be used in the exact manner described by the manufacturer.
  - Estimating a reference range  $N = 40\text{-}\underline{60}$ 
    - Used if the manufacturer's range is not adequate or if the use of the test not conform exactly to the manufacturer's intended use.
  - Establishing a reference range  $N \geq 120$ 
    - Non-FDA approved tests or if there will be significant changes to the use of the method.



# Reference Range Studies

- Transferring a reference range:
  - New reference range is calculated based on the systematic analytical differences between the two methods.
  - Can be done if the lab has previously established a reference range and is changing methodology
  - Acceptable, but not recommended method.
  - Should be verified by running at least 20 samples.
  - To reduce errors introduced by drift, transference calculations should be limited to one method change.



# Reference Range Studies

- “Divine judgment” of the Lab Director
  - Use only when all other options are unavailable.
  - May be needed for sub-population ranges.
  - Use published data from respected sources.



# Experiments to Validate?

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# Interference Studies

Materials in patient specimen that cause errors which are independent of analyte concentration

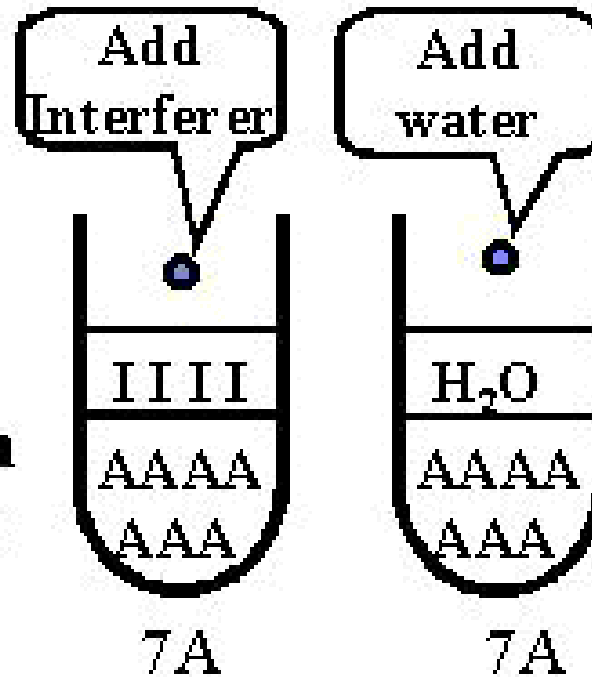
- Include substances commonly found in serum or plasma, such as:
  - Lipids (Lipemia)
  - Hemoglobin (Hemolysis)
  - Bilirubin (Icterus)
- Less common substances:
  - Drugs
- **Immunoassay Interferences:**
  - HAMA and other heterophile antibodies
  - Specific antibodies
  - Rheumatoid Factors
  - Non-specific binding of immunoglobulins (sticky serum, “anti-plastic”)
- Anticoagulants



# Interference Studies

## *The Interference Experiment*

**Prepare  
pairs of test  
samples**



**Measure A in  
both samples**

**Calculate  
difference**

$$7A - 7A = 0 \text{ bias}$$

From: [www.westgard.com](http://www.westgard.com)



# Interference Studies – “to do” list

- The interfering substance is “spiked” into a known sample (no analyte added)
- Added volume < 10%
- Run in duplicates
- Calculate interference (bias):

**Bias** = (sample + interference) - baseline sample

↓  
(sample + buffer/water)





# Interferences in Immunoassays

- Non-specific binding
  - High levels of immunoglobulins
  - Immune complexes
- Interfering antibodies
  - Rheumatoid factor
  - Specific antibodies to the analyte
  - Heterophile antibodies (antibodies to reagent non-human proteins)
- High concentrations of these types of substances may be difficult to obtain. Interference studies may require “mixing experiments”.



# Put Method On Line

- Write and test a procedure!
  - CLSI protocol (GP2)
  - Maintenance
  - Calibration
  - Control system
- Staff training
- Document Method Evaluation experiments according to appropriate regulations
- Start routine service
- Monitor performance



# Self Assessment Questions

1. Which of the following is a step in method validation?
  - a) **Error assessment**
  - b) Vendor consultation
  - c) FDA approval
  - d) Dissociative statistics

2. The lower limit of quantitation is defined as:
- a) The lowest number that will almost always have a non-zero result
  - b) The lowest concentration that can be distinguished from background
  - c) The lowest concentration that can be quantified reliably**
  - d) None of the above

3. The range of analyte where results are proportional to the true concentration of analyte in the sample without modification defines which of the following?
- a) Clinical reportable range
  - b) Precision
  - c) Analytical measurement range**
  - d) Accuracy

4. When evaluating a linear regression line ( $y = mx + b$ ), which of the following denotes the lowest level of proportional and constant bias?

a)  $y = .28x + .94$

b)  $y = 1.15x + .25$

**c)  $y = 1.05x - .04$**

d)  $y = .34x + 1.00$